

TOXICOLOGICAL SCREENING OF EXTRACTS FROM *CINNAMOMUM STENOPHYLLUM* ON *ARTEMIA SALINA* LEACH

TRIAGEM TOXICOLÓGICA DE EXTRATOS DE *CINNAMOMUM STENOPHYLLUM* FRENTE À *ARTEMIA SALINA* LEACH

Artur Danilo Novaes da Silva¹, Helder Carvalho Souza Lima Silva¹, Ricardo Prado Lyra¹, Bruno Lucêna de Lima¹, Daniela de Alencar Menezes¹, Gabriela Saraiva Dantas¹, Fabiana Lima Silva², Paulo Roberto, Hrihorowitsch Moreno³, Thárcia Kiara Beserra de Oliveira^{4,5}, Joelmir Lucena Veiga da Silva⁵

¹ Student at research group of integrative and complementary practices at Faculdade de Medicina de Olinda (FMO), ² Professor at the Institute of Health Sciences at Universidade Paulista (UNIP), ³ Professor at the Institute of Chemistry at Universidade Estadual de São Paulo (USP), ⁴ Professor at UNIFACISA, ⁵ Professor in the Research Group on Integrative and Complementary Practices at Faculdade de Medicina de Olinda (FMO)

ABSTRACT

Objective: To observe and compare the acute toxicity of the leaf (Csf) and stem extracts (Csc) from *Cinnamomum stenophyllum* against the brine shrimp *Artemia salina* Leach. **Methods:** Csf and Csc at concentrations of 1, 10, 100, and 1000 µg/mL were evaluated in acute toxicity assays on *A. salina* for 24 and 48 h, in triplicate. The number of dead nauplii was quantified, and the median lethal concentration (LC₅₀) values were calculated using non-linear regression. **Results:** Csf showed toxicity only at 1000 g/mL after 48 h ($p < 0.05$); therefore, the LC50 was not calculated. Csc exhibited toxicity only at 48 h of exposure (LC₅₀ = 8.7 ± 0.7 g/mL), indicating high toxicity (LC50 < 100 µg/mL). **Conclusion:** Csf and Csc from *C. stenophyllum* presented active metabolites that induced toxicity in *A. salina* under high exposure conditions. These metabolites are possibly different substances or are more concentrated in the stem. This is the first report of such findings in the literature.

Keywords: Medicinal plant; Toxicity; Vegetal extract

RESUMO

Objetivo: Observar e comparar a toxicidade aguda de extratos das folhas (Csf) e do caule (Csc) de *Cinnamomum stenophyllum* em *Artemia salina* Leach. **Métodos:** Os extratos Csf e Csc, nas concentrações de 1, 10, 100 e 1000 µg/mL, foram utilizados nos ensaios de toxicidade aguda utilizando o microcrustáceo *A. salina*, sob exposição de 24 e 48 horas, realizados em triplicata. O número de náuplios mortos foram quantificados e a CL50 foram calculadas por regressão não-linear. **Resultados:** Os extrato Csf apresentou toxicidade apenas com a concentração de 1000 g/mL em 48hs ($p < 0,05$), sendo assim, a CL50 não foi determinada. Já o extrato Csc foi tóxico apenas na maior exposição, de 48hs, mostrando CL50 de 8,7 0,7 g/mL, considerada uma alta toxicidade (CL50 < 100 g/mL). **Conclusão:** As folhas e caule de *C. stenophyllum* possuem metabólitos ativos que levam toxicidade a *A. salina* quando em alta exposição, os quais, provavelmente são substâncias diferentes ou estão mais concentradas no caule. Estes resultados são os primeiros na literatura para a espécie estudada.

Palavras-chave: Extrato vegetal; planta medicinal; toxicidade

INTRODUCTION

Traditional medicine and medicinal plants have been extensively used in developing countries, according to the norms for health maintenance¹.

According to the National Poison Information System, intoxication caused by medicinal plants

is the second leading cause of death by poisoning in humans. Several factors may contribute to this outcome, including limited knowledge regarding cultivation practices, incorrect plant identification, adverse reactions, drug interactions, dosage, and frequency of herbal medicine use².

The national guideline developed to support

and strengthen public health initiatives in Brazil is the National Policy for Integrative and Complementary Practices in the Unified Health System³. This policy initially encompassed the areas related to medicinal plants and phytotherapy, homeopathy, traditional Chinese medicine, acupuncture, and anthroposophic medicine. A more specific policy, the National Policy for Medicinal Plants and Herbal Medicines, was also implemented⁴.

The Lauraceae family is distributed across tropical and subtropical regions and comprises approximately 52 genera (between 2,500 and 3,500 species)⁵. In Brazil, this family is important and includes 23 genera and 420 species⁶, with many species producing aromatic oils and alkaloids used in perfumery and the pharmaceutical industry, such as *Cinnamomum camphora* (camphor) and *Aniba roseodora* (*pau-rosa*); the latter is a source of linalool, which is widely used in the cosmetics industry. Other species also produce edible fruits and culinary condiments, such as *Persea americana* (avocado), *Laurus nobilis* (laurel leaf), and *C. verum* (glycosylate)⁹.

As no studies reported the toxicological screening for *C. stenophyllum*, we aimed to evaluate and compare the acute toxicity of crude ethanolic extracts from the leaf (Csf) and stem (Csc) of this specimen on the microcrustacean *Artemia salina* Leach.

METHODS

Leaves and stems of *C. stenophyllum* were macerated in ethanol (95%). Extracts were obtained by solvent removal using a rotary evaporator at 60° C. The extracts were provided by the Instituto de Química at Universidade de São Paulo (USP). To prepare the solutions, extracts were solubilized in Cremophor (0.1%) and diluted in distilled water to a concentration of 2.5 mg/mL. Serial dilutions were performed to obtain the desired concentrations⁷. *C. stenophyllum* (Meisn.) Vattimo-Gil, known as “*canela-vassoura*”, is native to the Brazilian states of Minas Gerais, São Paulo, and Paraná^{6,8}. Extracts from species of the genus *Cinnamomum* have been reported to possess astringent, carminative, blood-purifying, digestive, antiseptic, antifungal, antiviral, antibacterial, antioxidant, anti-inflammatory, immunomodulatory, hypolipidemic, and hypoglyce-

mic properties⁹. Several chemical compounds have been reported in these species, including aldehydes, acetates, alcohols, terpenes, flavonoids, alkaloids, anthraquinones, coumarins, phenols, saponins, tannins, carboxylic acids, hydrocarbons, spathulenol, fatty acids, butanolides, lignans, steroids, prepenoids, and kaempferol.

The *A. salina* assay was used to determine the acute toxicity¹⁰. A quantity of 0.3 g of *A. salina* cysts was incubated in synthetic seawater for 24 to 36 h under artificial lighting at 22° C. After hatching, ten nauplii were transferred to test tubes containing the extract solutions (1, 10, 100, and 1000 µg/mL) or the control (saline). After 24 and 48 h, the number of live and dead larvae was quantified. Larvae were considered dead when no active movement was observed for approximately 20 seconds. The median lethal concentration (LC₅₀) for each extract was determined using non-linear regression analysis based on the number of viable nauplii at each concentration. The assay was conducted in triplicate for each tested concentration.

Results were expressed as mean ± standard error of the mean (X ± s.e.m.) and analyzed using the t-test; a p-value < 0.05 was considered statistically significant. Analyses were performed using GraphPad Prism software.

RESULTS AND DISCUSSION

Tests using *A. salina* are commonly applied to investigate the potential toxic activity of plant-derived products and for the preliminary screening and assessment of therapeutic safety. Due to its ease of maintenance in laboratory environments, *A. salina* has become a standard model for toxicity testing^{11,12}. The absence of cytotoxicity from the tested extracts in *A. salina* indicated that the plant material is well tolerated by this biological system.

Exposure to the Csf at different concentrations for 24 h did not affect *A. salina* viability compared with the control, indicating a lack of toxicity (Figure 1A). At 48 h of exposure, only the highest concentration (1000 g/mL) produced a statistically significant reduction in viability (Figure 1B). These results suggest that the stem of *C. stenophyllum* may contain active compounds that are not fully eliminated by the metabolism of *A. salina* when exposed to high concentrations, resulting in toxic effects.

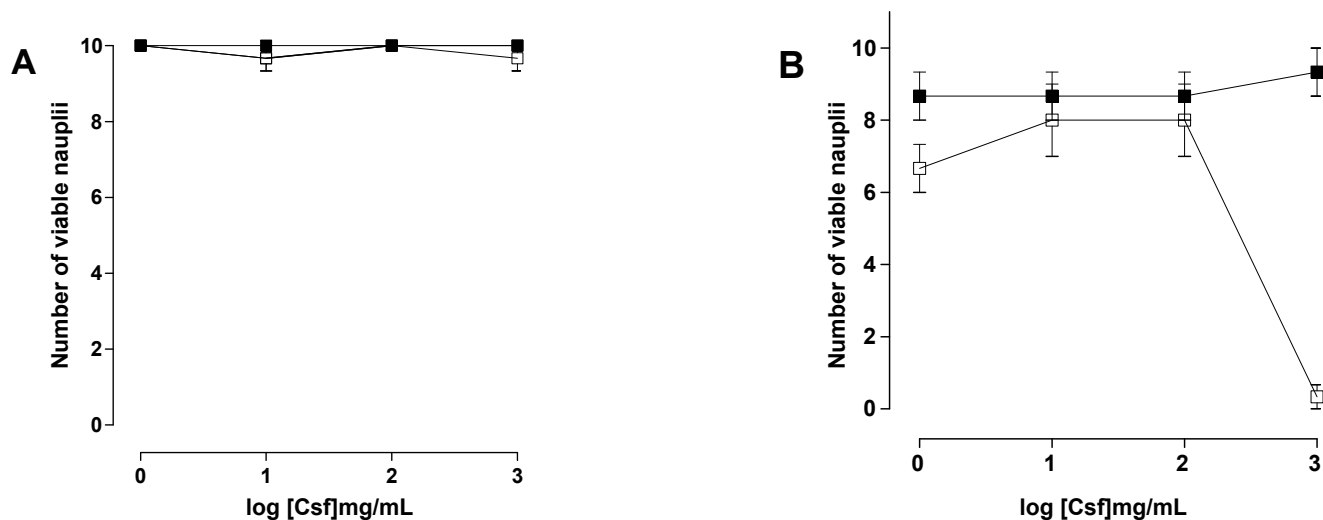


Figure 1. Viability of *Artemia salina* in the absence (■) or presence (□) of leaf extract (Csf) after 24 h (A) or 48 h (B) of exposure.

During the tests with the Csc (Figure 2), the mortality of *A. salina* was observed only after 48 h of exposure (Figure 2B); Csc produced an LC_{50} of 8.7 ± 0.7 g/mL, classified as highly toxic ($LC_{50} < 100$ g/mL)¹⁰. This toxicity was greater than that observed in the leaves and bark of *C. travancoricum*, *C. walaiwarens*, *C. wightii*, *C. verum*, *C. sulphuratum*, *C. riparium*, and *C. perrottetii*, and was performed in similar assays with *A. salina*¹³. The significant toxicity observed in the Csc suggests a potential for future cytotoxicity screening¹⁴. The greater toxicity of the Csc compared with the Csf of *C. stenophyllum* suggests that the active metabolites responsible for this effect may differ between plant parts or may occur at higher concentrations in the stem. Similar findings have been reported for bark extracts, which exhibited greater activity than the Csf of *C. travancoricum*, *C. walaiwarens*, *C. wightii*, *C. verum*, *C. sulphuratum*, *C. riparium*, and *C. perrottetii*¹³.

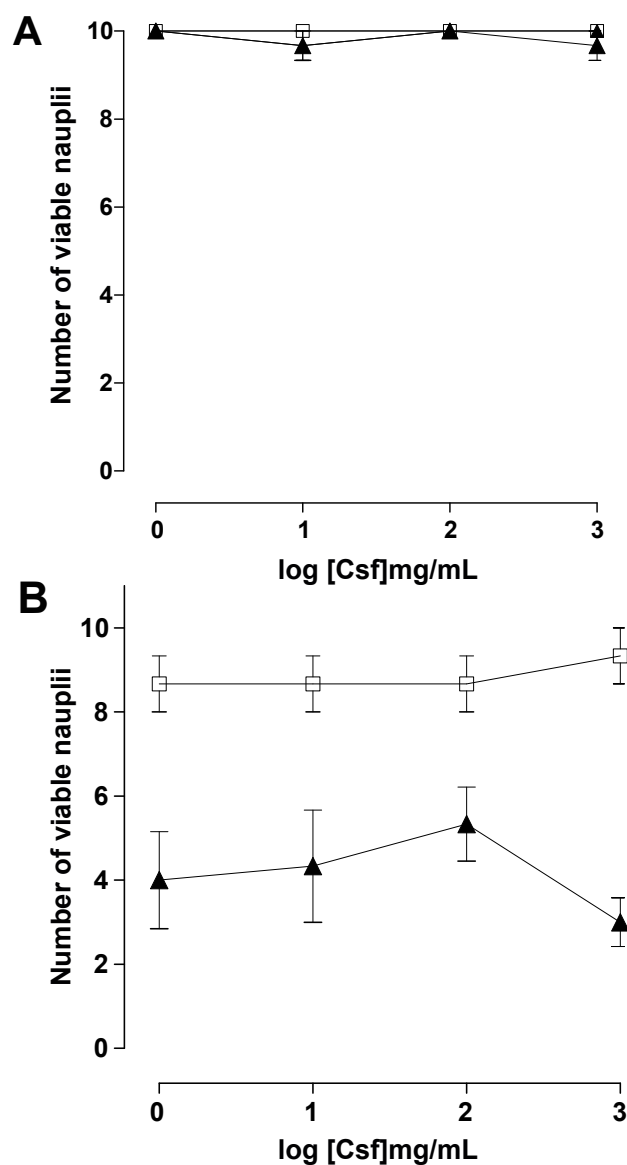


Figure 2. Viability of *Artemia salina* in the absence (□) or presence (▲) of stem extract (Csc) after 24 h (A) or 48 h (B) of exposure.

CONCLUSION

The leaves and stems of the *C. stenophyllum* contain active metabolites that cause toxicity to *A. salina* under prolonged exposure. The compounds responsible for this effect are probably different from those found in the leaves or are more concentrated in the stem. These are the first data reported for this plant specimen.

ACKNOWLEDGMENTS

We thank Rayane Rodrigues Angelo Viana for the technical support during the experiments conducted at the Laboratory of Functional Practices II of the Faculdade de Medicina de Olinda.

REFERENCES

1. Organização Das Nações Unidas Para A Educação, A Ciência E A Cultura (Unesco). Culture and Health: Orientation Texts: World Decade for Cultural Development 1988-1997, Document CLT/DEC/PRO. Paris, 1996;129.
2. Silveira PF, Bandeira MA, Arrais PSD. Farmacovigilância e reações adversas às plantas medicinais e fitoterápicos: uma realidade. Rev Bras Farmacogn 2008;18(4):618-26.
3. Ministério da Saúde. Portaria nº 971, de 03 de maio de 2006. Aprova a Política Nacional de Práticas Integrativas e Complementares (PNPIC BRASIL) no Sistema Único de Saúde. D.O.U. Poder Executivo, Brasília, 04 mai. 2006a.
4. Presidência da República. Decreto nº 5.813 de 22 de junho de 2006. Aprova a Política Nacional de Plantas Mediciniais e Fitoterápicos e dá outras providências. D.O.U. Poder Executivo, Brasília, 2006b.
5. Rohwer JG, Lauraceae. In: Kubitzki K, Rohwer JG, Bittrich V. (eds.). The families and genera of vascular plants 1993. Springer-Verlag, Berlin. Pp. 366-91.
6. Quinet A, Baitello JB, Moraes PLR, Lauraceae. In: Forzza RC, et al. (eds.). Lista de espécies da flora do Brasil. Disponível em: <<http://floradobrasil.jbrj.gov.br/2011/FB084232>>.
7. Santos S, Alves M. Flora da Usina São José, Igarassu, Pernambuco: Lauraceae. Rodriguésia 2012; 63(3):689-703.
8. Andrade TOD. Inventário e análise da arborização viária da Estância Turística de Campos do Jordão, SP. Monografia [graduação]. Piracicaba, SP: Universidade de São Paulo, 2002.
9. Kumar S, Kumari R, Mishra S. Pharmacological properties and their medicinal uses of Cinnamomum: a review. J Pharm and Pharmacol. 2019; 71:1735-61.
10. Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, Maclaughlin JL. Brine shrimp: a convenient general bioassay for active plant constituents. Plantas Mediciniais 1982; 45:31-34.
11. Amarante CB, Müller AH, Póvoa MM, Dolabela MF. Estudo fitoquímico biomonitorado pelos ensaios de toxicidade frente à *Artemia salina* e de atividade antiplasmódica do caule de aninga (*Montrichardia linifera*). Acta Amaz 2011;41(3):431-4.
12. Pimentel MF, Silva Junior FCG, Santaella ST, Lotufo LVC. O Uso de *Artemia* sp. como Organismo-Teste para Avaliação da Toxicidade das Águas Residuárias do Beneficiamento da Castanha de Caju Antes e Após Tratamento em Reator Biológico Experimental. J Braz Soc Ecotoxicol 2011; 6(1):15-22.
13. Maridass M. Evaluation of Brine Shrimp Lethality of Cinnamomum Species. Ethnobot Leaflets. 2008;12:772-5.
14. Barth EF, Pinto LS, Dileli P, Biavatti DC, Silva YL, Borolucci W, et al. Biological screening of extracts from leaf and stem bark of *Croton floribundus* Spreng. (Euphorbiaceae). Braz J Biol. 2018; 78(4):601-8.